INTERRELATIONSHIPS AMONG THE GUT, MITOCHONDRIAL FUNCTION, AND NEUROLOGICAL SEQUELAE

BY DANIEL A. ROSSIGNOL, MD, FAAFP

EVIDENCE OF A GUT-BRAIN CONNECTION

Overthe last century, a connection between gastrointestinal (GI) abnormalities and problems outside of the GI tract has become evident. For example, an association between GI problems and arthritis was described in 1910. Over time, a relationship between the GI tract and the brain (a gut-brain connection) also has emerged. As long ago as 1889, researchers reported "an exhaustional-confusional form of insanity proceeding from a dilated and over-filled colon." Colonic irrigation was commonly used in the late 1800s and early 1900s, with some investigators reporting that colon cleansing improved certain mental diseases.

Notwithstanding this history, it is only in the last decade or so that the gutbrain connection has become more widely acknowledged. Research in this area has greatly increased. While this article's overall focus is on the interaction between the gut and the brain, it highlights mitochondrial function as one of the critical bridges between these two body systems. I first examine some potential mechanisms of a gut-brain connection. Next, I discuss mitochondrial function in detail and assess how problems with mitochondrial function (mitochondrial dysfunction) can contribute to both GI abnormalities and neurological sequelae. In the context of abnormal GI function, I also review the potential adverse effects on mitochondrial function of bacterial imbalances in the GI tract and discuss how this can adversely affect the gut-brain connection. I conclude with a discussion of the potential role of hyperbaric oxygen therapy

(HBOT) in improving mitochondrial dysfunction as well as GI and brain function.

POTENTIAL MECHANISMS OF A GUT-BRAIN CONNECTION

Over time, a number of ideas have been developed to explain potential mechanisms of action for the gut-brain connection. One idea derives from evidence demonstrating that the central nervous system (CNS) and the GI tract share similar cells, including glial cells. In the GI tract, astrocyte-like glia are partly responsible for the proper functioning of the intestinal barrier and help to prevent larger food particles and other molecules from entering the circulatory system. Abnormalities in the GI glial cells may contribute to autoimmune diseases, enterocolitis, diabetes, irritable bowel syndrome (IBS), and inflammatory bowel disease (IBD).

The second body of evidence supporting a gut-brain connection comes from recent studies focusing on the bacteria in the GI tract. Whereas approximately 30,000 genes are found in the average human, more than 3 million genes from GI tract bacteria are present. The GI tract contains tenfold more bacteria (10^{14}) than the average number of cells (10^{13}) in a human body, and these bacteria serve important purposes. For example, the symbiotic relationship that exists between humans and their intestinal microbial flora is crucial for nutrient assimilation and important for the development of the innate immune system.



Exposure to "good" bacteria in the GI tract programs the immune system to more effectively fight infections. While it was already established that bacteria communicate with each other through a process known as quorum sensing, it is now apparent that the bacteria in the human GI tract also communicate with human cells through hormonal signals. This communication is important because bacteria and humans share metabolic pathways that are essential for health. 8

The third idea supporting a gut-brain connection comes from evidence that GI tract abnormalities may adversely affect brain function. Dysbiosis is the term used to refer to either an increase in the number of abnormal bacteria or a disruption in the type of bacteria in the GI tract. Although the idea that dysbiosis may contribute to abnormalities inside the GI tract (such as diarrhea and constipation) is fairly straightforward, it is increasingly apparent that dysbiosis also has effects outside the GI tract, including effects on brain function. Recent evidence has demonstrated that atypical levels and types of bacteria in the GI tract contribute to metabolic abnormalities reported in neurological and psychiatric conditions such as autism spectrum disorders (ASDs).9 The pathogenic metabolites produced by these bacteria in the GI tract may contribute to the brain dysfunction and metabolic problems that have been observed. As other examples, hydrogen sulfide produced in part by GI bacteria has been shown to play a role in blood pressure regulation, ¹⁰ and the unique makeup of the microbial community in the GI tract also may help determine a person's weight by influencing fat storage regulation. 11 Even a decade ago, the idea that the bacteria in the GI tract might influence blood pressure regulation or weight would have seemed untenable.

Fourth, dietary factors also point to a gut-brain connection. Perhaps nowhere is this more apparent than in the case of celiac disease, a condition

defined by intestinal damage resulting from gluten reactivity. Unfortunately, it has taken centuries for humans to realize that gluten exposure also can impair brain function in some people¹² and lead to conditions such as ataxia and schizophrenia. Interestingly, the prevalence of celiac disease is 3.5 times higher in children with ASD than in the general population.¹³ However, recent studies reveal that the general prevalence of celiac disease remains under-recognized.¹²

Exposure to foods other than glutencontaining foods may also impair brain function. For example, a one-month study reported disruptive behaviors in an 8-yearold boy with autism after exposure to a number of common foods. 14 Staff collected frequency data on behaviors such as object

throwing, scratching, biting, and screaming. The study included periods of a normal American diet, a fasting period, and a period during which individual foods were reintroduced one by one. During the latter phase, it was observed that mushrooms, dairy products, wheat, corn, tomatoes, and sugar all provoked behavioral problems. ¹⁴ Some children will manifest similar types of sensitivities. These reactions point to a connection between food that is ingested and effects in the brain, which then result in certain behavioral changes.

Exposure to cow's milk may also impair brain function in some people, including individuals with cerebral folate deficiency (CFD). CFD is a newly described neurodevelopmental disorder typified by low cerebrospinal fluid (CSF) levels of 5-methyltetrahydrofolate (5MTHF) (the metabolically active form of folic acid) in spite of normal systemic folic acid (folate) levels. 15 5MTHF is normally transported into the CNS through endocytosis by the cerebral folate receptor-alpha (FR α) in a process that is dependent on cellular energy. 15 The most common cause of CFD is circulating autoantibodies that bind to

the cerebral FR α and inhibit the transport of 5MTHF into the CSF. Lowered levels of 5MTHF in the CSF can lead to neurological abnormalities, including spastic paraplegia, cerebellar ataxia, dyskinesia, seizures, acquired microcephaly, and developmental regression that can occur as early as 4 months of age. ^{15,16} Central visual disturbances (optic atrophy and blindness) and hearing loss also have been described at later ages. ¹⁵ To date, seven studies have reported CFD in children with ASD, ^{15,17-22} and several studies have described CFD in Rett syndrome. ²³⁻²⁵ CFD has also been reported in individuals with mitochondrial disease, ^{26,27} perhaps because mitochondria are integral for providing the energy needed for the transport of 5MTHF into the brain. Treatment with oral folinic acid can lead to partial or complete recovery in some children. ^{15,17} In one such case, a 12-year-old girl with progressive spasticity, abnormal gait, and speech problems who had been diagnosed as paraplegic recovered from this condition after she was discovered to have CFD and was treated with oral folinic acid. ²⁸

Cow's milk contains soluble folate receptor antigen, which is 91% homologous to the FR α . Autoantibodies to the FR α cross-react with the soluble folate receptor antigen in cow's milk, which causes an increase in the circulating autoantibody concentration. Exposure to cow's milk has been shown to increase the concentration of the folate receptor autoantibody and lead to worsening of CFD symptoms, while elimination of cow's milk has been reported to lower the autoantibody concentration and improve CFD symptoms. ²⁰ Moreover, re-exposure to cow's milk after a period of being cow's milk-free substantially worsens the condition and increases the autoantibody concentration. ²⁰ These findings may help explain why some parents of children with ASD report improvements in their child on a cow's milk-free diet. ²⁹ Exposure to cow's milk also has been associated with constipation in children with ASD. ³⁰

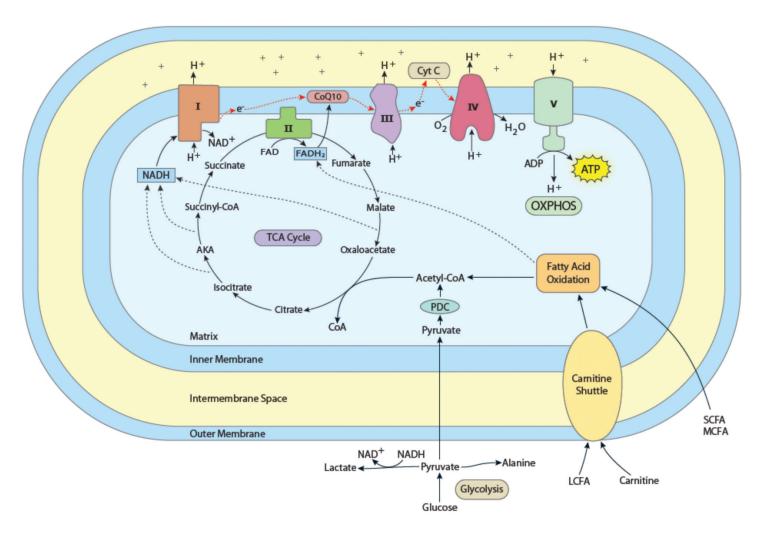
Besides the potential effect of dietary exposures on brain function, toxic byproducts produced by certain bacteria can also negatively affect brain function, especially in the face of liver dysfunction. For example, in individuals with hepatic encephalopathy, the intake of large amounts of protein can contribute to abnormal brain function. This occurs because protein is eventually broken down into ammonia, which is directly toxic to brain cells and can easily diffuse into the CNS from the bloodstream.31 High ammonia levels are associated with neuropsychological abnormalities in patients with hepatic encephalopathy.32 Because the mitochondria in the liver are responsible for the detoxification of ammonia, it is apparent that liver dysfunction contributes to high

ammonia. Ammonia is also produced by bacteria in the GI tract. High protein intake and dysbiosis in the face of liver dysfunction, therefore, can contribute to elevated ammonia levels and subsequent brain impairment. The important role of dysbiosis in this impairment is further illustrated by the finding that use of an antibiotic (rifaximin) reduces the risk of hepatic encephalopathy when compared with a placebo by eliminating ammonia-producing bacteria in the GI tract. 33

From this section, we can infer that mitochondrial function plays at least two important roles in maintaining a proper gut-brain connection. First, mitochondria help to detoxify certain toxins produced by GI bacteria that may otherwise adversely affect brain function. Second, mitochondria provide the energy needed to pump nutrients such as 5MTHF into the brain, a process that is impaired by autoantibodies that increase upon exposure to cow's milk. In the following sections, I review mitochondrial function and discuss how mitochondrial dysfunction can contribute to both GI abnormalities and neurological sequelae.

CFD is a newly described neurodevelopmental disorder typified by low cerebrospinal fluid (CSF) levels of 5-methyltetrahydrofolate (5MTHF) (the metabolically active form of folic acid) in spite of normal systemic folic acid (folate) levels.

Figure 1.



OVERVIEW OF MITOCHONDRIAL FUNCTION

Mitochondria are distinct cellular organelles that generate adenosine triphosphate (ATP) from adenosine diphosphate (ADP) by oxidizing glucose and fatty acids. ATP is the energy carrier in most mammalian cells. In the simplest terms, mitochondria are the powerhouses of the cell, generating energy from the breakdown of food. Figure 1 depicts a mitochondrion and shows the pathways involved when mitochondria break down food and use oxygen to create ATP (the energy source for the body, analogous to gasoline for a car). (For a more detailed review of mitochondrial function, see Haas and coauthors.³⁴)

As seen in Figure 1, the structure of the mitochondrion consists of outer and inner membranes, with a space (the intermembrane space) in between. The matrix is the innermost part of the mitochondria where many biochemical reactions occur, including the tricarboxylic acid (TCA) cycle (also known as the Krebs cycle or citric acid cycle). The inner mitochondrial membrane contains 5 complexes (known as complexes I through V) that make up the electron transport chain (ETC). On the bottom of the figure, you can see glucose, which is eventually broken down into pyruvate through the process of glycolysis. Pyruvate is then transported into the mitochondria and eventually is broken down into acetyl-CoA, which enters the TCA cycle.

Fatty acid metabolism is shown on the bottom right-hand corner of the figure. Short-chain fatty acids (SCFA) and medium-chain fatty acids (MCFA) can diffuse directly into the mitochondria, whereas long-chain fatty acids (LCFA) are transported into the mitochondria by attaching to carnitine, which shuttles these fatty acids across the inner and outer mitochondrial membranes. Once inside the mitochondria, the fatty acids, like pyruvate, are broken down

and converted into acetyl-CoA, which feeds into the TCA cycle. However, some of the electrons released from burning fatty acids (fatty acid oxidation) can feed into complex II through FADH2, bypassing complex I in the process (which may partially explain why a ketogenic diet, which involves a high intake of fats, might be helpful for treating mitochondrial dysfunction).³⁵

The three dotted lines with arrows coming off of the TCA cycle are electrons (negatively charged particles) that are transferred through NADH into complex I. Complex I then transfers these electrons to coenzyme Q10 (CoQ10) which, in turn, transfers the electrons to complex III. When the electrons pass through complex I, NADH is converted to NAD+. Hydrogen protons (positively charged hydrogen particles, H+) are pumped from the matrix (the innermost part of the mitochondria) through the inner membrane and into the intermembrane space, where they build up and form an electrochemical gradient. The electrons that passed to complex III are now transferred by cytochrome C (Cyt C) to complex IV. This process also pumps more hydrogen protons into the intermembrane space through complexes III and IV. During this process, oxygen is converted into water in complex IV. The hydrogen protons in the intermembrane space then diffuse back into the matrix through complex V (ATP synthase), and this generates ATP through a process known as oxidative phosphorylation.

MITOCHONDRIAL DYSFUNCTION

The concept of mitochondrial dysfunction is relatively new, and mitochondrial medicine is a rapidly evolving field of medicine. Mitochondrial disease, once thought uncommon, is now considered the most recognized cause of metabolic disease, ³⁶ with the minimum birth prevalence of an ETC defect estimated at 1 in 7634 individuals. ³⁷ If the ETC in the mitochondrion does not work properly (is blocked), metabolites begin to back up. Elevations can then occur in TCA cycle metabolites, fatty acids, and pyruvate. Moreover, pyruvate transportation can be slowed, and pyruvate can convert into elevated levels of lactate (also known as lactic acid) and alanine.

Mitochondrial disease has a broad phenotypic presentation: children with mitochondrial disease can have normal intelligence, mental retardation, or developmental delay. ³⁸ One recent study reported that approximately 5% of children with ASD have mitochondrial disease. ³⁹ It is important to note that illness or stress will generally place mitochondria under more stress and increase dysfunction. ³⁹ Thus, stressors such as dehydration, fever, and infection can lead to a functional decline and neurodegenerative regression in individuals with mitochondrial disease. ^{18,40}

To evaluate possible mitochondrial dysfunction, it is important to examine a patient's clinical history. Occasionally, there will be a family history of mitochondrial disease. Other clinical history that is often observed in mitochondrial dysfunction includes developmental regression (loss of previously acquired skills), seizures, fatigue or lethargy, ataxia (lack of coordination of muscle movements), motor delays, GI abnormalities (such as reflux, constipation, diarrhea, and inflammation), and cardiomyopathy (significant heart problems). Elevations in the various metabolites described previously are laboratory markers of mitochondrial dysfunction, making laboratory testing a helpful tool for identifying mitochondrial dysfunction. Generally, the higher the elevations and the more metabolites affected, the more likely it is that mitochondrial dysfunction exists.

The tests in question are typically covered by insurance and can be performed by any standard laboratory. The laboratory tests (ideally performed in the morning after fasting for 8-10 hours) may include those listed in Table 1. If the test results are abnormal, they may need to be repeated for confirmation. If the results are normal but mitochondrial dysfunction is still suspected, then repeating the tests when the child is sick or under stress may help unmask and identify mitochondrial dysfunction.

Table 1. Laboratory markers of mitochondrial dysfunction

- Lactate (lactic acid)
- Pyruvate
- Carnitine (free and total)
- Acylcarnitine panel (fatty acids attached to carnitine)
- Quantitative plasma amino acids (for measuring alanine and lysine)
- Ubiquinone (also known as CoQ10)
- Ammonia
- Creatine kinase (CK)
- AST (aspartate aminotransferase) and ALT (alanine aminotransferase)
- CO2 and glucose

MITOCHONDRIAL DYSFUNCTION, NEUROLOGICAL SEQUELAE, AND GI ABNORMALITIES

Since mitochondria are predominantly responsible for energy production, organs with the highest energy demand are most adversely affected by mitochondrial dysfunction. Low-energy cells (such as skin cells) have fewer mitochondria, while cells with higher energy needs contain more mitochondria. Cells that have high energy demands and, thus, many mitochondria include muscle, liver, brain, cerebrovascular endothelium, and GI cells. Neural synapses (areas of high energy consumption 11) are especially dependent on mitochondrial function. This helps explain how mitochondrial dysfunction can lead to impaired brain function. Mitochondria are concentrated in the dendritic and axonal termini, where they play an important role in ATP production, calcium homeostasis, and synaptic plasticity. Mitochondrial dysfunction can lead to reduced synaptic neurotransmitter release. Neurons that have high firing rates, such as GABAergic interneurons, may be the most adversely affected. Mitochondria also play an important role in cellular lipid metabolism, signaling, and repair. Mitochondria representations of the most adversely affected.

The ETC is the predominant source and the major target of reactive oxygen species (ROS). 48,49 ROS (also called free radicals) are molecules with an unpaired electron that can cause damage to cells and tissues by removing an electron from a surrounding compound or molecule. This damage is termed oxidative stress. Oxidative stress has been reported in many neurological disorders. 48,50 The ETC is protected from the damage caused by ROS by a mitochondrialspecific superoxide dismutase and by antioxidants such as glutathione (GSH). Under normal circumstances, GSH, the body's main antioxidant, can donate an electron to the free radical and quench the free radical before it can cause damage. Unfortunately, mitochondria lack the enzymes to synthesize GSH and, therefore, are dependent on cytosolic GSH production.^{51,52} When the GSH in mitochondria is depleted, cells are more vulnerable to oxidative stress and damage from ROS originating from the mitochondria.⁵³ Additionally, some factors can increase ROS production, including environmental toxins, infections, and autoimmune disease, which can directly and indirectly lead to impairments in ETC activity, 45,54,55 GSH depletion, 54 and activation of mitochondrial and non-mitochondrial-dependent biochemical cascades that result in programmed cell death (apoptosis).⁵⁶

Certain mammalian cells, such as neuronal and non-neuronal brain cells, are particularly vulnerable to oxidative stress. Moreover, the high rate of oxygen delivery and consumption in the brain provides the oxygen molecules necessary to generate ROS. The brain's ability to withstand oxidative stress is limited for the following reasons:

- a high content of substrates (such as polyunsaturated fatty acids) that are easily oxidized
- b relatively low levels of antioxidants, such as GSH and antioxidant enzymes
- c the endogenous generation of ROS via several specific reactions
- d the endogenous generation of nitric oxide (NO), a compound that readily transforms into reactive nitrogen species
- non-replicating cells that, if damaged, may become permanently dysfunctional or committed to apoptosis 54,56

Not surprisingly, mitochondrial dysfunction has been implicated in many neurological and psychiatric diseases, including neurodegenerative diseases

Since mitochondria are predominantly responsible for energy production, organs with the highest energy demand are most adversely affected by mitochondrial dysfunction.

The type of bacteria present in the GI tract may have a significant impact on the development of the brain and, eventually, on adult behavior.

Some bacteria in

the GI tract produce

metabolites that may

be potentially harmful.

such as Huntington's disease, Friedreich's ataxia, ⁴⁸ Parkinson's disease, and amyotrophic lateral sclerosis (ALS). ⁵⁷ Less obviously, mitochondrial dysfunction can also contribute to GI problems. For example, children with mitochondrial diseases are more likely to have GI abnormalities when compared with controls, ³⁹ and unexplained GI problems have been associated with mitochondrial disease. ⁵⁸ Constipation is a common symptom in children with mitochondrial diseases ³⁸ as is the more severe condition of obstipation (an inability to produce stool and gas). ⁵⁹ Given the high energy demands of both the GI tract and the cerebrovascular endothelium, it is apparent that mitochondrial dysfunction may contribute to barrier dysfunction in both the brain and GI tract. ³⁹

DYSBIOSIS, MITOCHONDRIAL DYSFUNCTION, AND NEUROLOGICAL SEQUELAE

The type of bacteria present in the GI tract may have a significant impact on the development of the brain and, eventually, on adult behavior. For example, the bacteria in the GI tract influence how the body uses vitamin B6, which then can affect the health of nerve cells. GI tract bacteria also have an influence on autoimmune diseases such as multiple sclerosis. A newly developed biochemical test that may help identify autism is based on the end products of GI bacterial metabolism.

Bacteria in the GI tract have been shown to influence anxiety. In one study, 60 mice lacking normal gut microflora were compared with mice with normal gut bacteria. Investigators looked at behavior, brain development, and brain chemistry. Illustrating one way in which the composition of the gut flora affect brain function, the animals lacking normal gut microflora actually had less anxiety as measured by some behavioral-related tests and displayed increased motor activity when compared with mice with normal gut flora. Two genes that may play a role in increasing anxiety—brain-derived neurotrophic factor (BDNF) and nerve growth factor-inducible clone A (NGF1-A)—were downregulated in the animals lacking the normal flora. In addition, the study demonstrated how the gut bacteria shape brain function by influencing brain cells to turn gene expression on or off, which in this instance affected the expression of almost 40 genes found in five different areas of the brain. These findings indicate that some brain-directed behaviors are influenced by the makeup of the GI flora. 60

Interestingly, some bacteria in the GI tract produce metabolites that may be otentially harmful. One example is propionic acid, an

potentially harmful. One example is propionic acid, an enteric short-chain fatty acid that is a fermentation end product of enteric bacteria. Propionic acid has been shown to inhibit mitochondrial fatty acid metabolism and function, ⁶¹⁻⁶⁵ contribute to seizure activity, ³⁹ and produce evidence of neuroinflammation, including reactive astrogliosis and activated microglia. ⁶¹ Clostridia, which are anaerobic, spore-forming Grampositive rod bacteria, are known to produce propionic acid, ⁶¹ and a derivative of propionic acid recovered in

the urine of individuals with ASD has been reported as a marker of clostridia. A recent rat model of ASD demonstrated that the administration of propionic acid induced mitochondrial dysfunction and led to brain, behavioral, and metabolic changes consistent with ASD, including clinical features such as repetitive behaviors, social interaction problems, hyperactivity, oxidative stress, lowered GSH levels, microglial activation, and altered carnitine levels. Furthermore, significantly elevated concentrations of clostridia in the GI tract have been reported in some children with ASD when compared with

controls ⁶⁷⁻⁷⁰ and in children with constipation when compared with controls. ⁷¹ Treatment of clostridia improves brain function in children with ASD ^{70,72} and has been shown to decrease hyperactivity and hypersensitivity as well as increase social interaction, eye contact, and vocalizations. ⁷⁰ Interestingly, clostridia levels increase with age and may lead to increased production of toxins that may affect liver function and play a role in cancer. ^{73,74}

My clinical experience confirms that a number of children with ASD have evidence of clostridia. Symptoms commonly associated with increased clostridia include hyperactivity, irritability, aggressiveness, increased self-stimulatory behavior, and obsessive behavior. Treatment of clostridia is often associated with a reduction in these behaviors, usually within several days to a week. In the context of clostridia in children with ASD, treatment with carnitine may be particularly helpful. This is because carnitine deficiency has been implicated in ASD, 75,76 some studies have reported improvements with the use of carnitine in ASD, 77-83 and carnitine may lower the toxicity 84 of propionic acid produced by clostridia. Carnitine has also been reported to improve mitochondrial function. 85

HBOT AND MITOCHONDRIAL DYSFUNCTION

Although treatments for mitochondrial dysfunction remain relatively limited, ³⁹ one treatment that has garnered interest in recent years is HBOT. HBOT involves inhaling up to 100% oxygen at a pressure greater than one atmosphere in a pressurized chamber. ⁸⁶ Since hypoxia is known to impair mitochondrial function, ⁸⁷ and because only approximately 0.3% of inhaled oxygen is ultimately delivered to mitochondria, ⁸⁸ increasing oxygen delivery to dysfunctional mitochondria through HBOT might aid in improving function. ^{89,90}

Both animal and human studies have examined the effects of HBOT on mitochondrial function. In a mouse model with an intrinsic impairment of mitochondrial complex IV, HBOT at 2 atmospheres "significantly ameliorate[d] mitochondrial dysfunction" and delayed the onset of motor neuron disease when compared with control mice. ⁸⁹ In other animal studies, HBOT increased the amount of work done by mitochondria, ⁹¹ improved mitochondrial function after brain injury, ⁹⁰ and prevented mitochondrial deterioration ⁹² when compared with room air pressure and 100% oxygen levels. HBOT has also been reported to increase sperm motility by augmenting mitochondrial oxidative

phosphorylation in fructolysis-inhibited sperm cells from rats. Fructose is the sugar used by sperm for energy production. In rats, HBOT prevented apoptosis and improved neurological recovery after cerebral ischemia by opening mitochondrial ATP-sensitive potassium channels. In another animal model, hypoxia and ischemia led to diminished ATP and phosphocreatine production; the addition of HBOT restored these levels to near normal and increased energy utilization when compared with room air oxygen and pressure levels.

In an animal model, HBOT was recently shown to activate mitochondrial DNA transcription and replication and increase the biogenesis of mitochondria in the brain. HBOT could be used to increase the production of mitochondria in humans. Although biogenesis has not yet been proven to occur in humans, I have had two patients with severe mitochondrial disease and abnormal mitochondrial function (as measured by muscle biopsy) who have improved clinically with HBOT at 1.3 to 1.5 atmospheres and who now have normal mitochondrial function (again as measured by muscle biopsy).

In a recent human study of 69 patients with severe traumatic brain injury, HBOT at 1.5 atmospheres and 100% oxygen significantly increased brain oxygen levels, increased cerebral blood flow, and decreased CSF lactate levels. It also improved brain metabolism and mitochondrial function when compared with both room air treatment and 100% oxygen given at normobaric pressure. 97

HBOT AND GI FUNCTION

Because HBOT possesses potent anti-inflammatory properties, 98,99 it may be useful in ameliorating inflammatory conditions of the GI tract. Several published animal models of IBD have demonstrated that HBOT can significantly lower inflammation in the GI tract and improve IBD. 99-104 Other studies have reported improvements using HBOT for Crohn's disease 105-108 and ulcerative colitis. 109-111 In addition, HBOT can kill clostridial species, because these bacteria are anaerobic. 112 In my clinical experience, children with evidence of clostridia often have significant improvements in GI function (especially in diarrhea) with the use of HBOT.

HBOT AND BRAIN FUNCTION

HBOT may help improve brain function in certain conditions. In animal models, HBOT improves learning and memory. 113 In healthy young adults, the addition of 100% oxygen when compared with room air significantly enhances memory,¹¹⁴ cognitive performance (including word recall and reaction time),115 attention, and picture recognition.116 Several studies have

shown improvements in traumatic or chronic brain injury with HBOT. 117-120 In a recent study of 16 individuals who had traumatic brain injury, individuals exhibited significant improvements with the use of HBOT at 1.5 atmospheres and 100% oxygen in their neurological exam, IQ, memory, post-traumatic stress symptoms, depression, and anxiety; they also displayed objective improvements in brain perfusion.¹²¹ Studies also have reported behavioral improvements in children with ASD using HBOT at 1.3 to 1.5 atmospheres. 122-126 HBOT may bring about improvements even in conditions where permanent brain problems are thought to be present, including cerebral palsy 127,128 and fetal alcohol syndrome. 129

CONCLUSIONS

The evidence for a gut-brain connection has become stronger over time. Abnormalities in the GI tract, including dysbiosis, and abnormalities caused by intake of gluten and cow's milk proteins may contribute to abnormal brain function. Mitochondria play an important role in the gut-brain connection, and abnormalities in mitochondrial function are found in many neurological and psychiatric disorders. Mitochondrial dysfunction can lead to GI abnormalities and brain dysfunction. Ultimately, mitochondrial dysfunction can have adverse effects on the gut-brain connection. Treatment of mitochondrial dysfunction with modalities such as carnitine and HBOT may be beneficial in maintaining and/or improving the gut-brain connection. Additional studies examining the gut-brain connection in neurological and psychiatric disorders are warranted.

REFERENCES

- Marshall HW. Arthritis of gastrointestinal origin, its diagnosis and treatment. JAMA. 1910;55(22):1871
- Moyer HN. Insanity proceeding from the colon. JAMA. 1889;13(7):230.
- Marshall HK, Thompson CE. Colon irrigation in the treatment of mental disease. New Engl J Med. 1932;207(10):454-7.
- Savidge TC, Sofroniew MV, Neunlist M. Starring roles for astroglia in barrier pathologies of gut and brain. Lab Invest. 2007;87(8):731-6.
- Clarke TB, Davis KM, Lysenko ES, Zhou AY, Yu Y, Weiser JN. Recognition of peptidoglycan from the microbiota by Nod1 enhances systemic innate immunity. Nat Med. 2010;16(2):228-31.
- Hughes DT, Sperandio V. Inter-kingdom signalling: communication between bacteria and their hosts. Nat Rev Microbiol. 2008;6(2):111-20.
- Sperandio V, Torres AG, Jarvis B, Nataro JP, Kaper JB. Bacteria-host communication: the language of hormones. Proc Natl Acad Sci USA. 2003;100(15):8951-6.
- Nicholson JK, Holmes E, Lindon JC, Wilson ID. The challenges of modeling mammalian biocomplexity. Nat Biotechnol. 2004;22(10):1268-74.
- Yap IK, Angley M, Veselkov KA, Holmes E, Lindon JC, Nicholson JK. Urinary metabolic phenotyping differentiates children with autism from their unaffected siblings and age-matched controls. J Proteome Res. 2010;9(6):2996-3004.
- Yang G, Wu L, Jiang B, Yang W, Qi J, Cao K et al. H2S as a physiologic vasorelaxant: hypertension in mice with deletion of cystathionine gamma-lyase. Science. 2008;322(5901):587-90.
- Bäckhed F, Ding H, Wang T, Hooper LV, Koh GY, Nagy A et al. The gut microbiota as an environmental factor that regulates fat storage. Proc Natl Acad Sci USA. 2004;101(44):15718-23.
- 12. Hadjivassiliou M, Grunewald RA, Davies-Jones GA. Gluten sensitivity as a neurological illness. J Neurol Neurosurg Psychiatry. 2002;72(5):560-3.
- 13. Barcia G, Posar A, Santucci M, Parmeggiani A. Autism and coeliac disease. J Autism Dev Disord. 2008;38(2):407-8.
- O'Banion D, Armstrong B, Cummings RA, Stange J. Disruptive behavior: a dietary approach. J Autism Child Schizophr. 1978;8(3):325-37.

- 15. Ramaekers VT, Blau N. Cerebral folate deficiency. Dev Med Child Neurol. 2004;46(12):843-51.
- Ramaekers VT, Hausler M, Opladen T, Heimann G, Blau N. Psychomotor retardation, spastic paraplegia, cerebellar ataxia and dyskinesia associated with low 5-methyltetrahydrofolate in cerebrospinal fluid: a novel neurometabolic condition responding to folinic acid substitution. Neuropediatrics. 2002;33(6):301-8.
- Ramaekers VT, Blau N, Sequeira JM, Nassogne MC, Quadros EV. Folate receptor autoimmunity and cerebral folate deficiency in low-functioning autism with neurological deficits. Neuropediatrics. 2007;38(6):276-81
- Shoffner J, Hyams L, Langley GN, Cossette S, Mylacraine L, Dale J et al. Fever plus mitochondrial disease could be risk factors for autistic regression. J Child Neurol. 2010;25(4):429-34.
- Ramaekers VT, Rothenberg SP, Sequeira JM, Opladen T, Blau N, Quadros EV et al. Autoantibodies to folate receptors in the cerebral folate deficiency syndrome. N Engl J Med. 2005;352(19):1985-91.
- Ramaekers VT, Sequeira JM, Blau N, Quadros EV. A milk-free diet downregulates folate receptor autoimmunity in cerebral folate deficiency syndrome. Dev Med Child Neurol. 2008;50(5):346-52
- Moretti P, Peters SU, Del Gaudio D, Sahoo T, Hyland K, Bottiglieri T et al. Brief report: autistic symptoms developmental regression, mental retardation, epilepsy, and dyskinesias in CNS folate deficiency. J Autism Dev Disord. 2008;38(6):1170-7.
- 22. Moretti P, Sahoo T, Hyland K, Bottiglieri T, Peters S, del Gaudio D et al. Cerebral folate deficiency with developmental delay, autism, and response to folinic acid. Neurology. 2005;64(6):1088-90.
- Ramaekers VT, Hansen SI, Holm J, Opladen T, Senderek J, Hausler M et al. Reduced folate transport to the CNS in female Rett patients. Neurology. 2003;61(4):506-15.
- 24. Ramaekers VT, Sequeira JM, Artuch R, Blau N, Temudo T, Ormazabal A et al. Folate receptor autoantibodies and spinal fluid 5-methyltetrahydrofolate deficiency in Rett syndrome. Neuropediatrics. 2007;38(4):179-83.
- 25. Perez-Duenas B, Ormazabal A, Toma C, Torrico B, Cormand B, Serrano M et al. Cerebral folate deficiency syndromes in childhood: clinical, analytical, and etiologic aspects. Arch Neurol. 2011;68(5):615-21.

- 26. Ramaekers VT, Weis J, Sequeira JM, Quadros EV, Blau N. Mitochondrial complex I encephalomyopathy and cerebral 5-methyltetrahydrofolate deficiency. Neuropediatrics. 2007;38(4):184-7.
- 27. Garcia-Cazorla A. Quadros EV. Nascimento A. Garcia-Silva MT, Briones P, Montoya J et al. Mitochondrial diseases associated with cerebral foliate deficiency. Neurology. 2008;70(16):1360-2.
- 28. Hansen FJ, Blau N. Cerebral folate deficiency: lifechanging supplementation with folinic acid. Mol Genet Metab. 2005;84(4):371-3.
- Whiteley P, Haracopos D, Knivsberg AM, Reichelt KL, Parlar S, Jacobsen J et al. The ScanBrit randomised, controlled, single-blind study of a gluten- and casein-free dietary intervention for children with autism spectrum disorders. Nutr Neurosci. 2010;13(2):87-100.
- 30. Afzal N, Murch S, Thirrupathy K, Berger L, Fagbemi A, Heuschkel R. Constipation with acquired megarectum in children with autism. Pediatrics. 2003;112(4):939-42.
- 31. Cagnon L, Braissant O. Hyperammonemia-induced toxicity for the developing central nervous system. Brain Res Rev. 2007;56(1):183-97.
- 32. Weissenborn K, Ennen JC, Schomerus H, Ruckert N, Hecker H. Neuropsychological characterization of hepatic encephalopathy. J Hepatol. 2001;34(5):768-73
- 33. Bass NM, Mullen KD, Sanyal A, Poordad F, Neff G, Leevy CB et al. Rifaximin treatment in hepatic encephalopathy. N Engl J Med. 2010;362(12):1071-81.
- 34. Haas RH, Parikh S, Falk MJ, Saneto RP, Wolf NI, Darin N et al. Mitochondrial disease: a practical approach for primary care physicians. Pediatrics. 2007;120(6):1326-33.
- 35. Kang HC, Lee YM, Kim HD, Lee JS, Slama A. Safe and effective use of the ketogenic diet in children with epilepsy and mitochondrial respiratory chain complex defects. Epilepsia. 2007;48(1):82-8.
- **36.** Zeviani M, Bertagnolio B, Uziel G. Neurological presentations of mitochondrial diseases. *J Inherit Metab* Dis. 1996;19(4):504-20.
- 37. Skladal D, Halliday J, Thorburn DR. Minimum birth prevalence of mitochondrial respiratory chain disorders in children. Brain. 2003;126(Pt 8):1905-12.
- Nissenkorn A, Zeharia A, Lev D, Watemberg N, Fattal-Valevski A, Barash V et al. Neurologic presentations of mitochondrial disorders. J Child Neurol. 2000;15(1):44-8.

REFERENCES

- Rossignol DA, Frye RE. Mitochondrial dysfunction in autism spectrum disorders: a systematic review and meta-analysis. Mol Psychiatry. 2011 Jan 25.
- Edmonds JL, Kirse DJ, Kearns D, Deutsch R, Spruijt L, Naviaux RK. The otolaryngological manifestations of mitochondrial disease and the risk of neurodegeneration with infection. Arch Otolaryngol Head Neck Surg. 2002; 128(4):355-62.
- Ames A, 3rd. CNS energy metabolism as related to function. Brain Res Brain Res Rev. 2000;34(1-2):42-68.
- Mattson MP, Liu D. Energetics and oxidative stress in synaptic plasticity and neurodegenerative disorders. Neuromolecular Med. 2002;2(2):215-31.
- Chen H, Chan DC. Mitochondrial dynamics-fusion, fission, movement, and mitophagy-in neurodegenerative diseases. Hum Mol Genet. 2009;18(R2):R169-76.
- 44. Li Z, Okamoto K, Hayashi Y, Sheng M. The importance of dendritic mitochondria in the morphogenesis and plasticity of spines and synapses. Cell. 2004;119(6):873-87.
- 45. Anderson MP, Hooker BS, Herbert MR. Bridging from cells to cognition in autism pathophysiology: biological pathways to defective brain function and plasticity. Am J Biochem Biotech. 2008;4(2):167-76.
- Marin-Garcia J, Goldenthal MJ. Heart mitochondria signaling pathways: appraisal of an emerging field. J Mol Med. 2004;82(9):565-78.
- Goldenthal MJ, Marin-Garcia J. Mitochondrial signaling pathways: a receiver/integrator organelle. Mol Cell Biochem. 2004;262(1-2):1-16.
- Trushina E, McMurray CT. Oxidative stress and mitochondrial dysfunction in neurodegenerative diseases. Neuroscience. 2007;145(4):1233-48.
- Fernandez-Checa JC, Garcia-Ruiz C, Colell A, Morales A, Mari M, Miranda M et al. Oxidative stress: role of mitochondria and protection by glutathione. Biofactors. 1998;8(1-2):7-11.
- Chauhan A, Chauhan V. Oxidative stress in autism. Pathophysiology. 2006;13(3):171-81.
- Enns GM. The contribution of mitochondria to common disorders. Mol Genet Metab. 2003;80(1-2):11-26.
- James SJ, Rose S, Melnyk S, Jernigan S, Blossom S, Pavliv
 O et al. Cellular and mitochondrial glutathione redox
 imbalance in lymphoblastoid cells derived from children with
 autism. FASEB J. 2009;23(8):2374-83.
- Fernandez-Checa JC, Kaplowitz N, Garcia-Ruiz C, Colell A, Miranda M, Mari M et al. GSH transport in mitochondria: defense against TNF-induced oxidative stress and alcoholinduced defect. Am J Physiol. 1997;273(1 Pt 1):G7-17.
- Calabrese V, Lodi R, Tonon C, D'Agata V, Sapienza M, Scapagnini G et al. Oxidative stress, mitochondrial dysfunction and cellular stress response in Friedreich's ataxia. J Neurol Sci. 2005;233(1-2):145-62.
- Munnich A, Rustin P. Clinical spectrum and diagnosis of mitochondrial disorders. Am J Med Genet. 2001;106(1):4-17.
- Roberts RA, Laskin DL, Smith CV, Robertson FM, Allen EM, Doorn JA et al. Nitrative and oxidative stress in toxicology and disease. Toxicol Sci. 2009;112(1):4-16.
- Martin LJ. Mitochondriopathy in Parkinson disease and amyotrophic lateral sclerosis. J Neuropathol Exp Neurol. 2006;65(12):1103-10.
- Verma A, Piccoli DA, Bonilla E, Berry GT, DiMauro S, Moraes CT. A novel mitochondrial G8313A mutation associated with prominent initial gastrointestinal symptoms and progressive encephaloneuropathy. Pediatr Res. 1997;42[4]:448-54.
- Skladal D, Sudmeier C, Konstantopoulou V, Stockler-Ipsiroglu S, Plecko-Startinig B, Bernert G et al. The clinical spectrum of mitochondrial disease in 75 pediatric patients. Clin Pediatr (Phila). 2003;42(8):703-10.
- Heijtz RD, Wang S, Anuar F, Qian Y, Bjorkholm B, Samuelsson A et al. Normal gut microbiota modulates brain development and behavior. Proc Natl Acad Sci USA 2011;108(7):3047-52.
- 61. MacFabe DF, Cain DP, Rodriguez-Capote K, Franklin AE, Holfman JE, Boon F et al. Neurobiological effects of introventricular propionic acid in rats: possible role of short chain fatty acids on the pathogenesis and characteristics of autism spectrum disorders. Behav Brain Res. 2007;176(1):149-69.

- 62. MacFabe DF, Rodríguez-Capote K, Hoffman JE, Franklin AE, Mohammad-Asef Y, Taylor AR et al. A novel rodent model of autism: intraventricular infusions of propionic acid increase locomotor activity and induce neuroinflammation and oxidative stress in discrete regions of adult rat brain. Am J Biochem Biotech. 2008;4(2):146-66.
- Schwab MA, Sauer SW, Okun JG, Nijtmans LG, Rodenburg RJ, van den Heuvel LP et al. Secondary mitochondrial dysfunction in propionic aciduria: a pathogenic role for endogenous mitochondrial toxins. Biochem J. 2006;398(1):107-12.
- 64. Shultz SR, MacFabe DF, Ossenkopp KP, Scratch S, Whelan J, Taylor R et al. Intracerebroventricular injection of propionic acid, an enteric bacterial metabolic end-product, impairs social behavior in the rat: implications for an animal model of autism. Neuropharmacology. 2008;54(6):901-11.
- 65. Thomas RH, Foley KA, Mepham JR, Tichenoff LJ, Possmayer F, MacFabe DF. Altered brain phospholipid and acylcarnitine profiles in propionic acid infused rodents: further development of a potential model of autism spectrum disorders. J Neurochem. 2010;113(2):515-29.
- 66. Shaw W. Increased urinary excretion of a 3-(3-hydroxyphenyl)-3-hydroxypropionic acid (HPHPA), an abnormal phenylalanine metabolite of Clostridia spp. in the gastrointestinal tract, in urine samples from patients with autism and schizophrenia. Nutr Neurosci. 2010;13(3):135-43.
- Parracho HM, Bingham MO, Gibson GR, McCartney AL. Differences between the gut microflora of children with autistic spectrum disorders and that of healthy children. J Med Microbiol. 2005;54(Pt 10):987-91.
- Finegold SM, Molitoris D, Song Y, Liu C, Vaisanen ML, Bolte E et al. Gastrointestinal microflora studies in late-onset autism. Clin Infect Dis. 2002;35(Suppl 1):S6-S16.
- Song Y, Liu C, Finegold SM. Real-time PCR quantitation of clostridia in feces of autistic children. Appl Environ Microbiol. 2004;70(11):6459-65.
- Bolte ER. Autism and Clostridium tetani. Med Hypotheses. 1998;51(2):133-44.
- Zoppi G, Cinquetti M, Luciano A, Benini A, Muner A, Bertazzoni Minelli E. The intestinal ecosystem in chronic functional constipation. Acta Paediatr. 1998;87(8):836-41.
- Sandler RH, Finegold SM, Bolte ER, Buchanan CP, Maxwell AP, Vaisanen ML et al. Short-term benefit from oral vancomycin treatment of regressive-onset autism. J Child Neurol. 2000;15(7):429-35.
- Mallett AK, Rowland IR, Farthing MJ. Dietary modification of intestinal bacterial enzyme activities-potential formation of toxic agents in the gut. Scand J Gastroenterol Suppl. 1987;129:251-7.
- Goldin BR. Intestinal microflora: metabolism of drugs and carcinogens. Ann Med. 1990;22(1):43-8.
- GA, El-Gamal HA, El-Wakkad ASE, El-Shorbagy OE, Hamza MM. Polyunsaturated fatty acids, carniline and lactate as biological markers of brain energy in autistic children. Int J Child Neuropsychiatry. 2005;2(2):179-88.
- Filipek PA, Juranek J, Nguyen MT, Cummings C, Gargus JJ. Relative carnitine deficiency in autism. J Autism Dev Disord. 2004;34(6):615-23.
- Gargus JJ, Imtiaz F. Mitochondrial energy-deficient endophenotype in autism. Am J Biochem Biotech. 2008;4(2):198-207.
- Poling JS, Frye RE, Shoffner J, Zimmerman AW. Developmental regression and mitochondrial dysfunction in a child with autism. J Child Neurol. 2006;21(2):170-2.
- Pastural E, Ritchie S, Lu Y, Jin W, Kavianpour A, Khine Su-Myat K et al. Novel plasma phospholipid biomarkers of autism: mitochondrial dysfunction as a putative causative mechanism. Prostaglandins Leukot Essent Fatty Acids. 2009;81(4):253-64.
- Geier DA, Kern JK, Davis G, King PG, Adams JB, Young JL et al. A prospective double-blind, randomized clinical trial of levocarnitine to treat autism spectrum disorders. Med Sci Monit. 2011;17(6):P115-23.
- Gargus JJ, Lerner MA. Familial autism with primary carnitine deficiency, sudden death, hypotonia and hypochromic anemia. Am J Human Gen. 1997;61:A98.
- Filipek PA, Juranek J, Smith M, Mays LZ, Ramos ER, Bocian M et al. Mitochondrial dysfunction in autistic patients with 15q inverted duplication. Ann Neurol. 2003;53(6):801-4.

- Ezugha H, Goldenthal M, Valencia I, Anderson CE, Legido A, Marks H. 5q14.3 deletion manifesting as mitochondrial disease and autism: case report. J Child Neurol. 2010;25(10):1232-5.
- Lucke T, Perez-Cerda C, Baumgartner M, Fowler B, Sander S, Sasse M et al. Propionic acidemia: unusual course with late onset and fatal outcome. Metabolism. 2004;53(6):809-10.
- Frye RE, Rossignol DA. Mitochondrial dysfunction can connect the diverse medical symptoms associated with autism spectrum disorders. *Pediatr Res.* 2011;69(5 Pt 2):41 R-47R.
- 86. Feldmeier JJ, Chairman and Editor. Hyperbaric Oxygen 2003: Indications and Results: The Hyperbaric Oxygen Therapy Committee Report. Kensington, MD: Undersea and Hyperbaric Medicine Society, 2003.
- Magalhaes J, Ascensao A, Soares JM, Ferreira R, Neuparth MJ, Marques F et al. Acute and severe hypobaric hypoxia increases oxidative stress and impairs mitochondrial function in mouse skeletal muscle. J Appl Physiol. 2005;99(4)b1247-53.
- Lane N. Oxygen: The Molecule that Made the World. New York, NY: Oxford University Press, 2003.
- Dave KR, Prado R, Busto R, Raval AP, Bradley WG, Torbati D et al. Hyperbaric oxygen therapy protects against mitochondrial dysfunction and delays onset of motor neuron disease in Wobbler mice. Neuroscience. 2003;120(1):113-20.
- Daugherty WP, Levasseur JE, Sun D, Rockswold GL, Bullock MR. Effects of hyperbaric oxygen therapy on cerebral oxygenation and mitochondrial function following moderate lateral fluid-percussion injury in rats. J Neurosurg. 2004;101(3):499-504.
- Boveris A, Chance B. The mitochondrial generation of hydrogen peroxide. General properties and effect of hyperbaric oxygen. Biochem J. 1973;134(3):707-16.
- 92. Gosalvez M, Castillo Olivares J, De Miguel E, Blanco M, Figuera D. Mitochondrial respiration and oxidative phosphorylation during hypothermic hyperbaric hepatic preservation. J Surg Res. 1973;15(5):313-8.
- Bar-Sagie D, Mayevsky A, Bartoov B. Effects of hyperbaric oxygenation on spermatozoan motility driven by mitochondrial respiration. J Appl Physiol. 1981;50(3):531-7.
- Lou M, Chen Y, Ding M, Eschenfelder CC, Deuschl G. Involvement of the mitochondrial ATP-sensitive potassium channel in the neuroprotective effect of hyperbaric oxygenation after cerebral ischemia. Brain Res Bull. 2006;69(2):109-16.
- Calvert JW, Zhang JH. Oxygen treatment restores energy status following experimental neonatal hypoxiaischemia. Pediatr Crit Care Med. 2007;8(2):165-73.
- Gutsaeva DR, Suliman HB, Carraway MS, Demchenko IT, Piantadosi CA. Oxygen-induced mitochondrial biogenesis in the rat hippocampus. Neuroscience. 2006;137(2):493-504.
- 97. Rockswold SB, Rockswold GL, Zaun DA, Zhang X, Cerra CE, Bergman TA et al. A prospective, randomized clinical trial to compare the effect of hyperbaric to normobaric hyperoxia on cerebral metabolism, intracranial pressure, and oxygen toxicity in severe traumatic brain injury. J Neurosurg. 2010;112(5):1080-94.
- Granowitz EV, Skulsky EJ, Benson RM, Wright J, Garb JL, Cohen ER et al. Exposure to increased pressure or hyperbaric oxygen suppresses interferon-gamma secretion in whole blood cultures of healthy humans. Undersea Hyperb Med. 2002;29(3):216-25.
- Gorgulu S, Yagci G, Kaymakcioglu N, Ozkara M, Kurt B, Ozcan A et al. Hyperbaric oxygen enhances the efficiency of 5-aminosalicylic acid in acetic acid-induced colitis in rats. Dig Dis Sci. 2006;51(3):480-7.
- 100. Akin ML, Gulluoglu BM, Uluutku H, Erenoglu C, Elbuken E, Yildirim S et al. Hyperbaric oxygen improves healing in experimental rat colitis. Undersea Hyperb Med. 2002;29(4):279-85.
- 101. Atug O, Hamzaoglu H, Tahan V, Alican I, Kurtkaya O, Elbuken E et al. Hyperbaric oxygen therapy is as effective as dexamethasone in the treatment of TNBS-E-induced experimental colitis. Dig Dis Sci. 2008;53(2):481-5.

- 102. Gulec B, Yasar M, Yildiz S, Oter S, Akay C, Deveci S et al. Effect of hyperbaric oxygen on experimental acute distal colitis. Physiol Res. 2004;53(5):493-9.
- 103. Guven A, Gundogdu G, Uysal B, Cermik H, Kul M, Demirbag S et al. Hyperbaric oxygen therapy reduces the severity of necrotizing enterocolitis in a neonatal rat model. J Pediatr Surg. 2009;44(3):534-40.
- **104.** Rachmilewitz D, Karmeli F, Okon E, Rubenstein I, Better OS. Hyperbaric oxygen: a novel modality to ameliorate experimental colitis. *Gut.* 1998;43(4):512-8.
- 105. Brady CE 3rd, Cooley BJ, Davis JC. Healing of severe perineal and cutaneous Crohn's disease with hyperbaric oxygen. Gastroenterology. 1989;97(3):756-60.
- Colombel JF, Mathieu D, Bouault JM, Lesage X, Zavadil P, Quandalle P et al. Hyperbaric oxygenation in severe perineal Crohn's disease. Dis Colon Rectum. 1995;38(6):609-14
- 107. Lavy A, Weisz G, Adir Y, Ramon Y, Melamed Y, Eidelman S. Hyperbaric oxygen for perianal Crohn's disease. J Clin Gastroenterol. 1994;19(3):202-5.
- 108. Takeshima F, Makiyama K, Doi T. Hyperbaric oxygen as adjunct therapy for Crohn's intractable enteric ulcer. Am J Gastroenterol. 1999;94(11):3374-5.
- 109. Kuroki K, Masuda A, Uehara H, Kuroki A. A new treatment for toxic megacolon. Lancet. 1998;352(9130):782.
- Buchman AL, Fife C, Torres C, Smith L, Aristizibal J. Hyperbaric oxygen therapy for severe ulcerative colitis. J Clin Gastroenterol. 2001;33(4):337-9.
- Gurbuz AK, Elbuken E, Yazgan Y, Yildiz S. A different therapeutic approach in patients with severe ulcerative colitis: hyperbaric oxygen treatment. South Med J. 2003;96(6):632-3.
- 112. Ml, Erenoglu C, Dal A, Erdemoglu A, Elbuken E, Batkin A. Hyperbaric oxygen prevents bacterial translocation in rats with obstructive jaundice. Dig Dis Sci. 2001;46(8):1657-62.

- 113. Harch PG, Kriedt C, Van Meter KW, Sutherland RJ. Hyperbaric oxygen therapy improves spatial learning and memory in a rat model of chronic traumatic brain injury. Brain Res. 2007;1174:120-9.
- 114. Moss MC, Scholey AB. Oxygen administration enhances memory formation in healthy young adults. Psychopharmacology (Berl). 1996;124(3):255-60.
- 115. Scholey AB, Moss MC, Neave N, Wesnes K. Cognitive performance, hyperoxia, and heart rate following oxygen administration in healthy young adults. *Physiol Behav*. 1999;67(5):783-9.
- 116. Moss MC, Scholey AB, Wesnes K. Oxygen administration selectively enhances cognitive performance in healthy young adults: a placebocontrolled double-blind crossover study. Psychopharmacology (Berl). 1998;138(1):27-33.
- 117. Hardy P, Johnston KM, De Beaumont L, Montgomery DL, Lecomte JM, Soucy JP et al. Pilot case study of the therapeutic potential of hyperbaric oxygen therapy on chronic brain injury. J Neurol Sci. 2007;253(1-2):94-105
- 118. Harch PG, Fogarty EF, Staab PK, Van Meter K. Low pressure hyperbaric oxygen therapy and SPECT brain imaging in the treatment of blast-induced chronic traumatic brain injury (post-concussion syndrome) and post traumatic stress disorder: a case report. Cases J. 2009;2:6538.
- Golden Z, Golden CJ, Neubauer RA. Improving neuropsychological function after chronic brain injury with hyperbaric oxygen. Disabil Rehabil. 2006;28(22):1379-86.
- 120. Golden Zl, Neubauer R, Golden CJ, Greene L, Marsh J, Mleko A. Improvement in cerebral metabolism in chronic brain injury after hyperbaric oxygen therapy. Int J Neurosci. 2002;112(2):119-31.

- 121. Harch P, Andrews SR, Fogarty E, Amen DG, Pezzullo JC, Lucarini J et al. A Phase I study of low-pressure hyperbaric oxygen therapy for blast-induced post-concussion syndrome and post-traumatic stress disorder. J Neurotrauma. 2011 Nov 22.
- Rossignol DA, Rossignol LW. Hyperbaric oxygen therapy may improve symptoms in autistic children. Med Hypotheses. 2006;67(2):216-28.
- 123. Rossignol DA, Rossignol LW, James SJ, Melnyk S, Mumper E. The effects of hyperbaric oxygen therapy on oxidative stress, inflammation, and symptoms in children with autism: an open-label pilot study. BMC Pediatr. 2007;7(1):36.
- 124. Rossignol DA, Rossignol LW, Smith S, Schneider C, Logerquist S, Usman A et al. Hyperbaric treatment for children with autism: a multicenter, randomized, doubleblind, controlled trial. BMC Pediatr. 2009;9:21.
- 125. Chungpaibulpatana J, Sumpatanarax T, Thadakul N, Chantharatreerat C, Konkaew M, Aroonlimsawas M. Hyperbaric oxygen therapy in Thai autistic children. J Med Assoc Thai. 2008;91(8):1232-8.
- 126. Bent S, Bertoglio K, Ashwood P, Nemeth E, Hendren RL. Brief report: hyperbaric oxygen therapy (HBOT) in children with autism spectrum disorder: a clinical trial. J Autism Dev Disord. 2011.
- 127. Marois P, Vanasse M. Hyperbaric oxygen therapy and cerebral palsy. Dev Med Child Neurol. 2003;45(9):646-7; author reply 7-8.
- 128. Sénéchal C, Larivée S, Richard E, Marois P. Hyperbaric oxygenation therapy in the treatment of cerebral palsy: a review and comparison to currently accepted therapies. Journal of American Physicians and Surgeons. 2007;12(4):109-13.
- 129. Stoller KP. Quantification of neurocognitive changes before, during, and after hyperboric oxygen therapy in a case of fetal alcohol syndrome. *Pediatrics*. 2005;116(4):e586-91.

NeuroProtek®



NeuroProtek® and our new formulation, NeuroProtek® LP are all natural, flavonoid based, nutritional supplements. Both formulas, based on the published scientific research of Prof. Theoharides, PhD, MD., contain an exclusive combination of three pure, plant based flavonoids: Luteolin, Quercetin and Rutin. Research has shown these ingredients inhibit oxidative stress and inflammation, while reducing gut and brain barrier disruption. Unique to Algonot's formulas is micro-filtered, low acidity, Olive Kernel Oil (OKO). OKO contains many antioxidant components and is instrumental in helping the body absorb and deliver flavonoid powders. NeuroProtek® & NeuroProtek® LP are free of the following allergens: artificial colors or dyes, flavors or sweeteners, corn, eggs, fish, heavy metals, milly(asein, peanuts, preservatives, salt, shellfish, soy, starch, sugar, tree nuts, wheat/gluten and yeast.

1-800-254-6668 1-941-346-5304 feedback@algonot.com

Luteolin® - Flavonoid



Protects Nerve Degeneration Prevents Blood-Brain Barrier Disruption Blocks Oxidative Stress Modulates Immune System Reduces Inflammation Inhibits Mast Cell Activation

Quercetin - Flavonoid



Inhibits the Release of IL-6 & TNF Reduces Inflammation Blocks Oxidative Stress Inhibits Mast Cell Activation

Rutin - Flavonoid



The Natural Glycoside of Quercetin Liberates Quercetin in the Gut Reduces Oxidative Stress Reduces Gut-Blood Barrier Disruption

Olive Kernel Oil



Increases Absorption of Flavonoids Contains Anti-Oxidant Components Protects Mucosal Surfaces Micro-Filtered, Low-Acidity

